

Going Wide with Targeted Proteomics: The Rise of Unhole-y Data

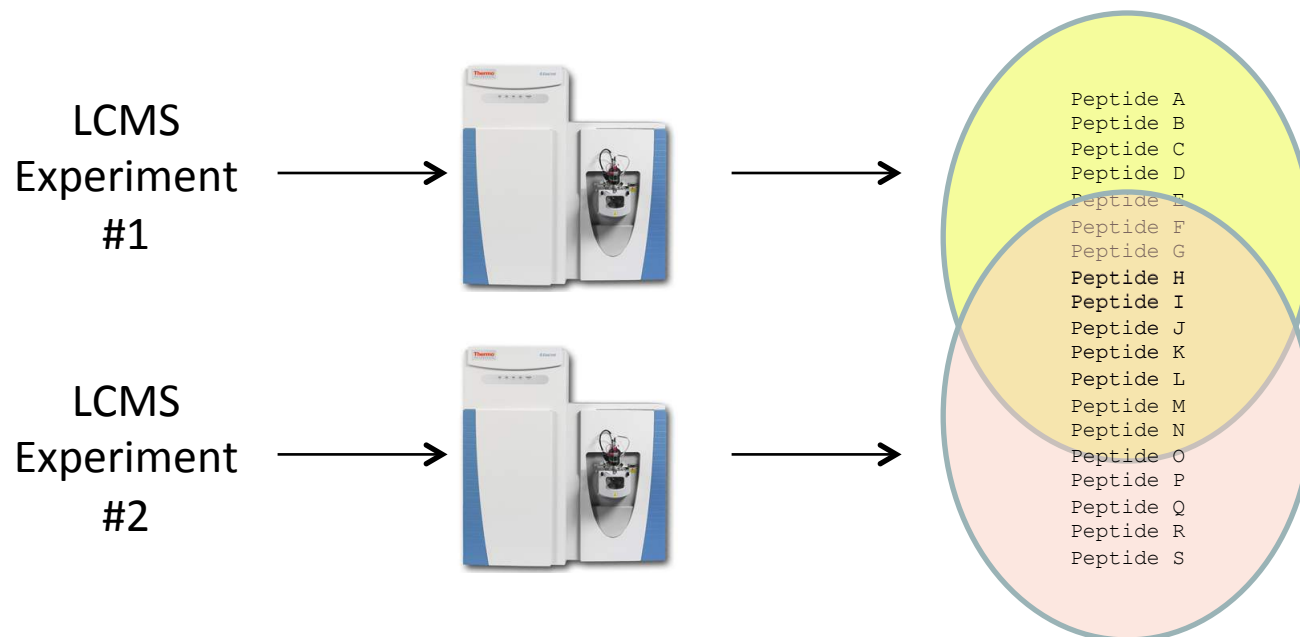
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The Broad Institute

LINCS Data Forum March 2013

Shotgun Proteomics Experiments are Stochastic

- Instrumentation inherently undersamples population
 - Speed
 - Sensitivity
 - Dynamic Range



Data Shapes in Proteomics

1. Long and skinny

Peptides/proteins	1		2	
	1	2	1	2
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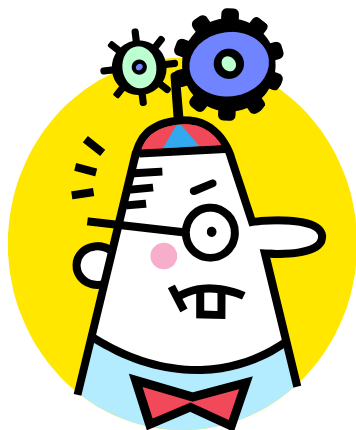
2. Hole-y

[illegible]

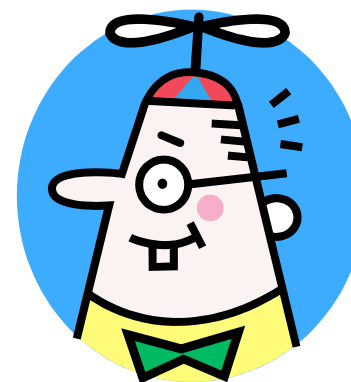
3. Short and wide

[illegible]

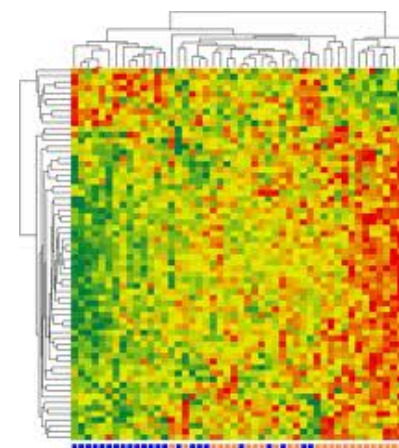
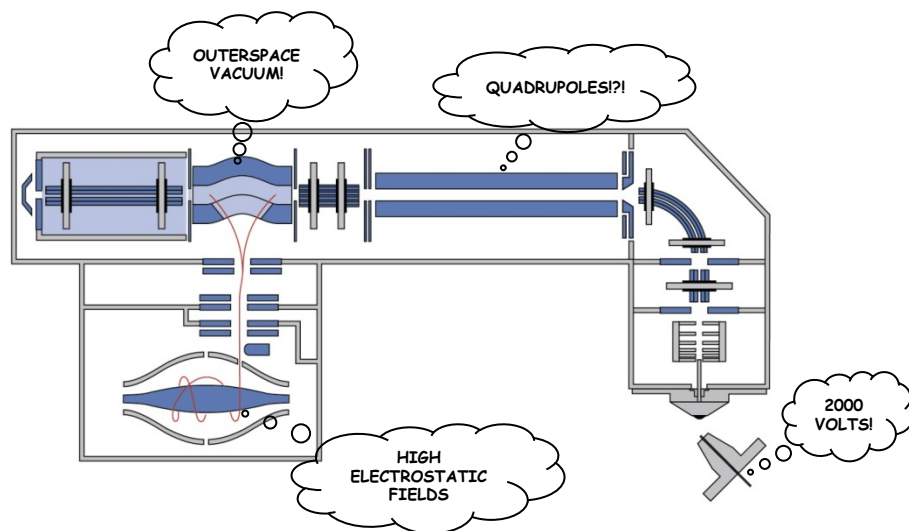
It's hard to be hole-y



Instrument Jockeys



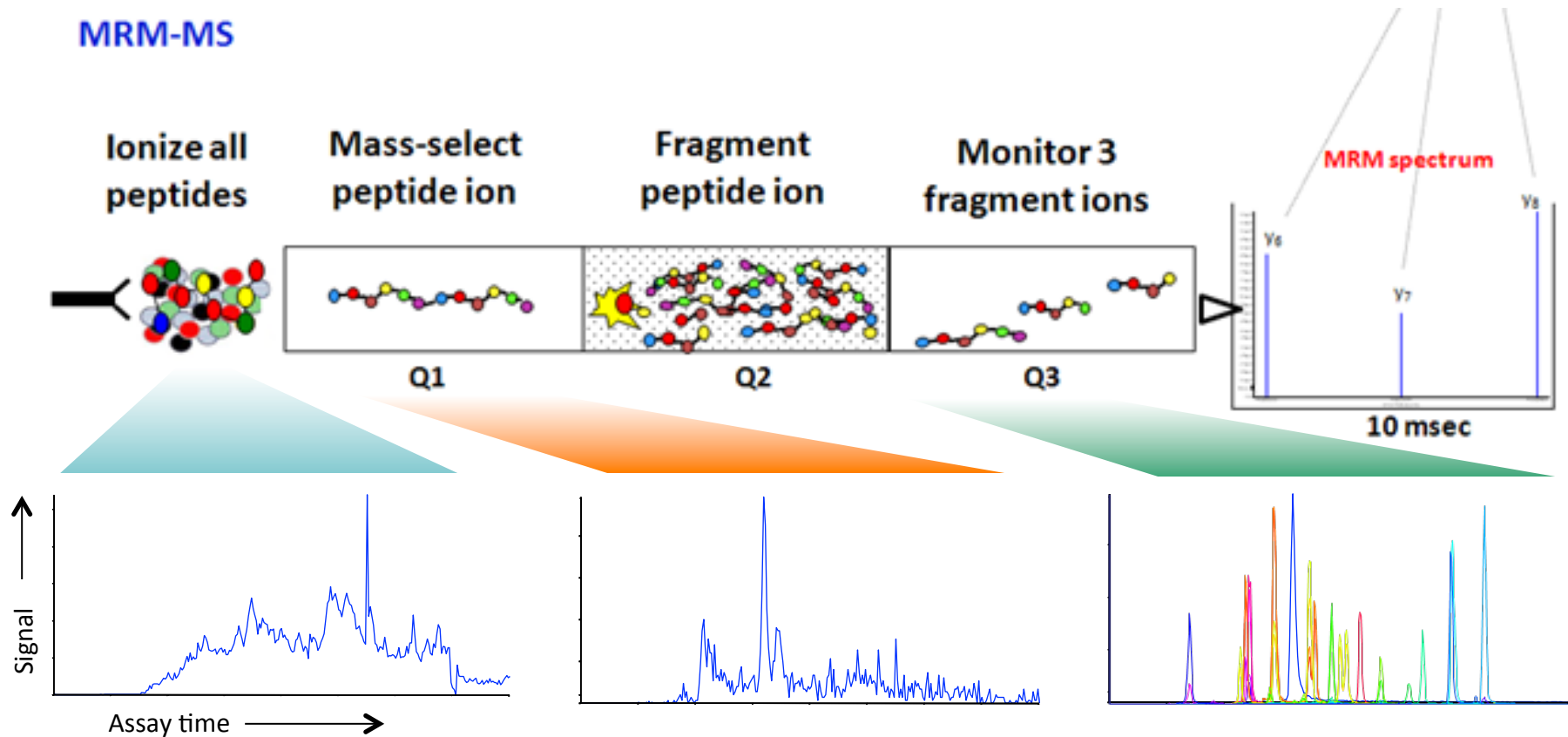
Data Geniuses



Solution: Targeted Proteomics

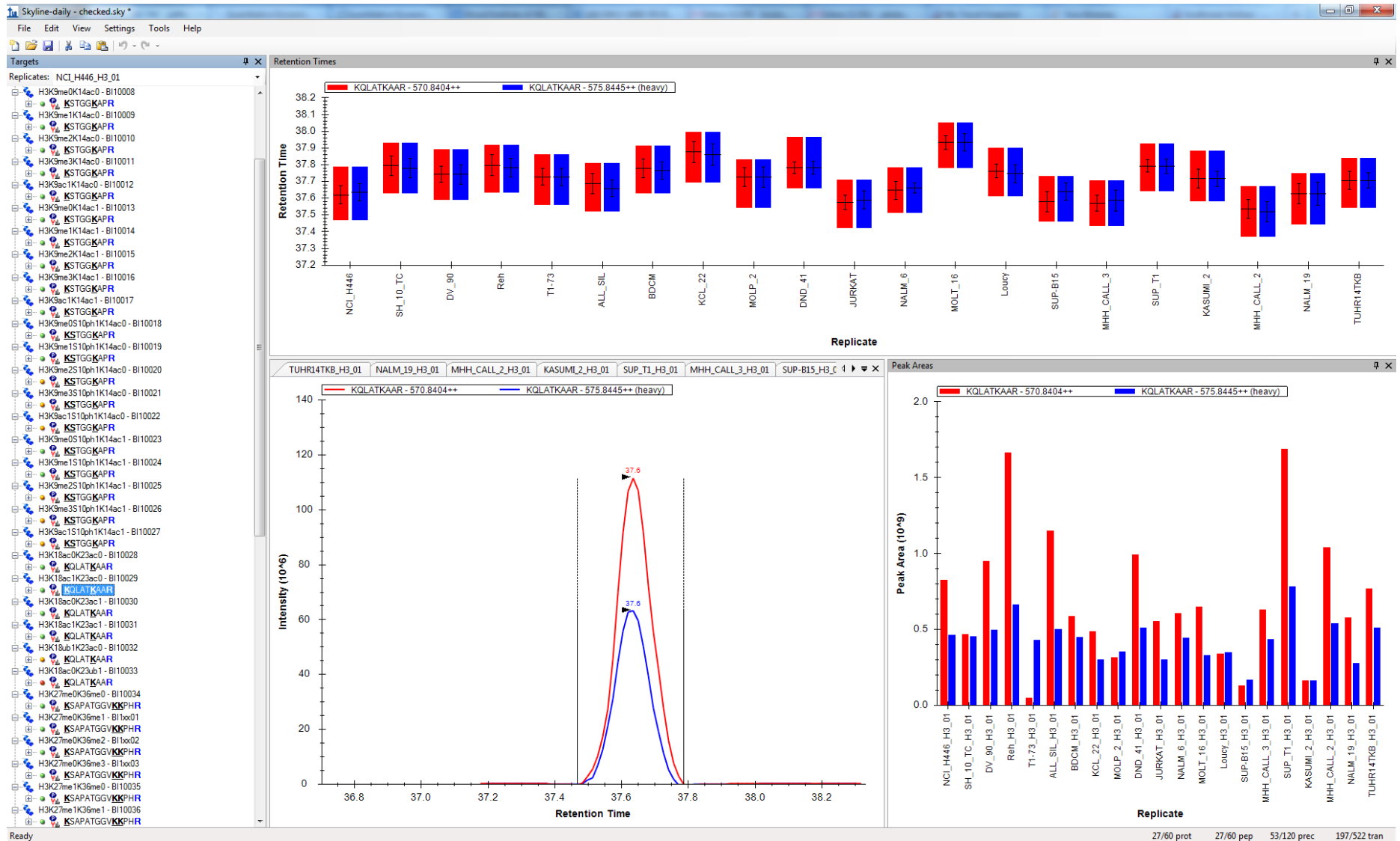
- Limit proteomic assay to finite number of meaningful analytes
 - Multiplex ~ 100-200
- Introduce internal standards for guaranteed observation of analytes
- Internal standards = better quantification
- When holoproteomics is not an option

Targeted Proteomics Implementation

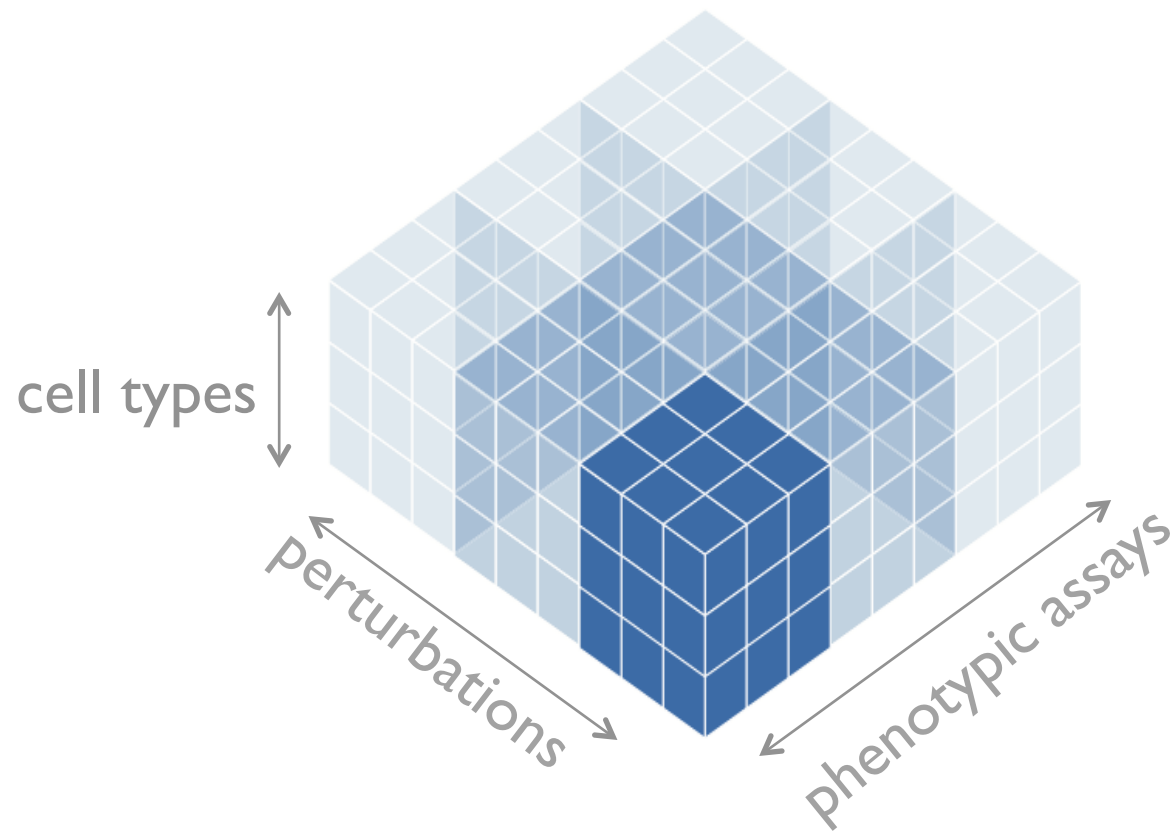


- Just need to know m/z , r.t., and fragment m/z s
- Generally cheaper and faster than shotgun proteomics

Target Proteomics Quantification



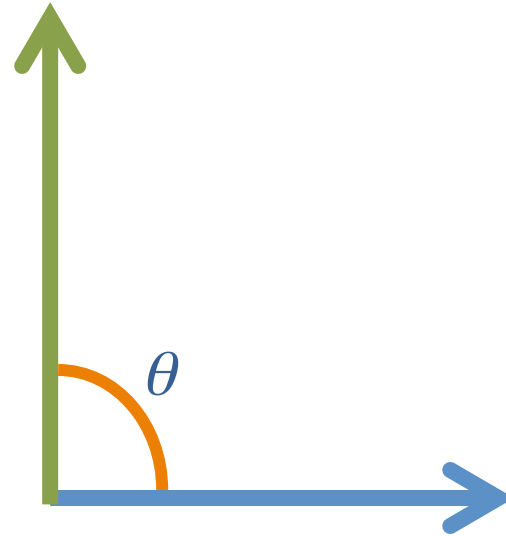
Targeted Proteomics and LINCS



- Desirable to have “complete” data in every assay

Targeted Phosphoproteomics Application: P100

Phospho-signaling

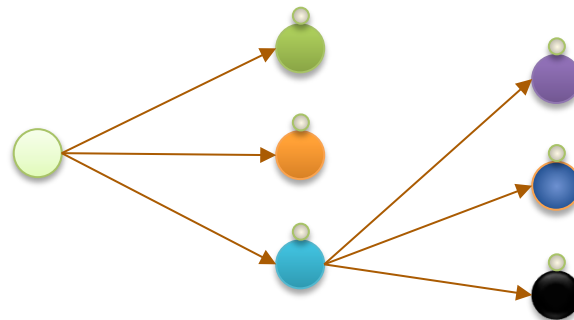


Gene Expression

- θ may be large
- Phosphosignaling is inaccessible through expression profiling
- Phosphosignaling can occur on different time scales than gene expression

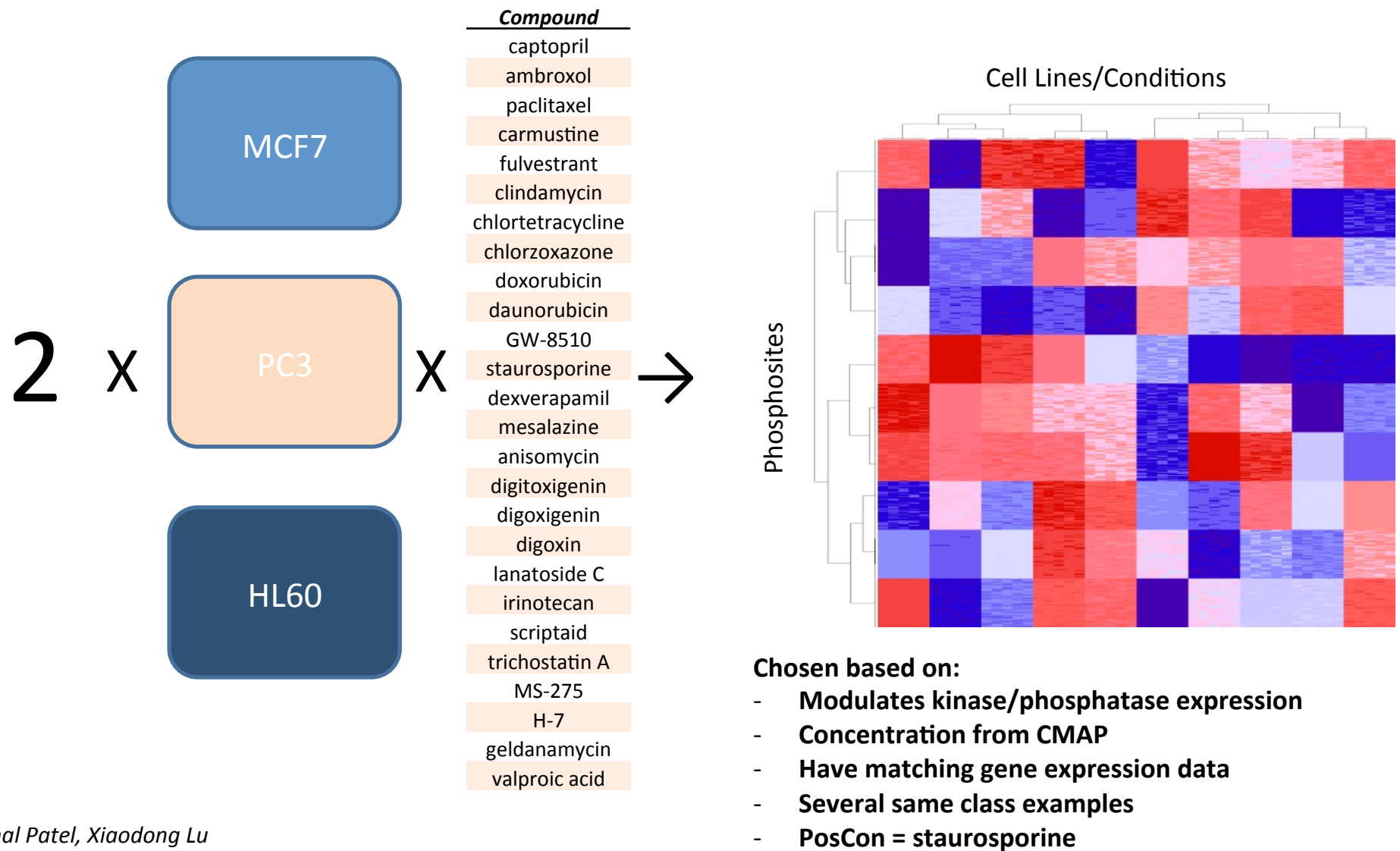
Reduced Representation Signaling Set: P100

- We are building a phosphosignaling analog of the L1000 assay – the P100
- Phosphosignaling is likely coordinated
 - Kinase-substrate relationships tend to be 1-to-many
 - *A priori* expectation of coordinate regulation of sites



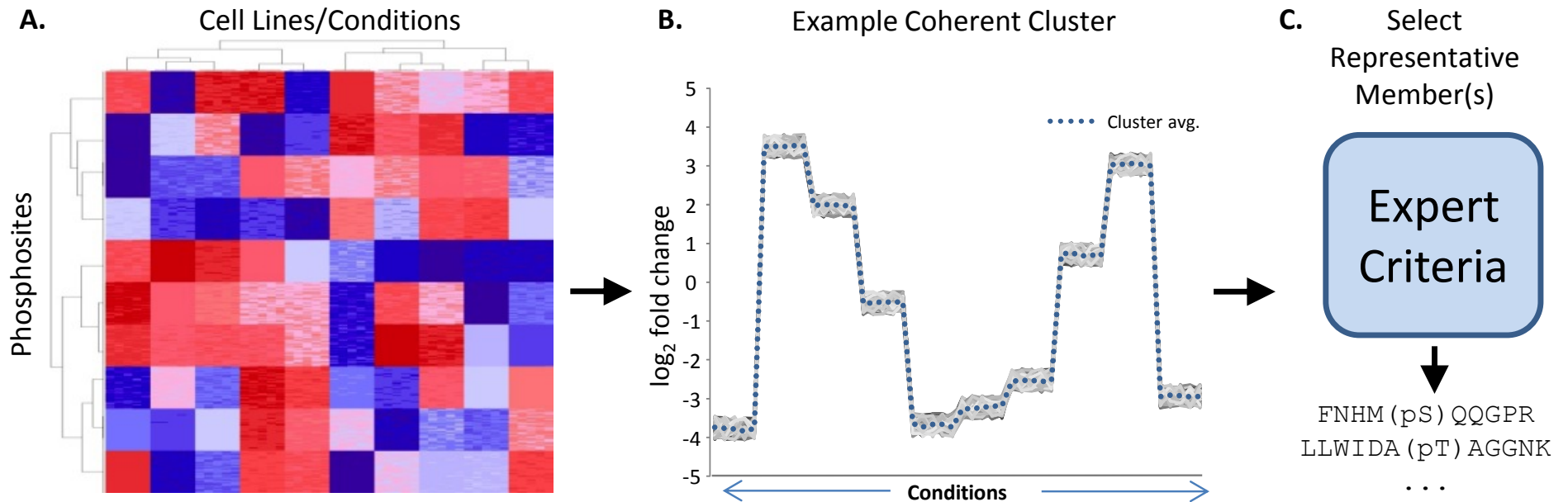
- Don't need to monitor every phosphosite in the assay
 - Amenable to L1000-like reductionist approaches

Generation of source data from LINCS Lines and Compounds



Jinal Patel, Xiaodong Lu

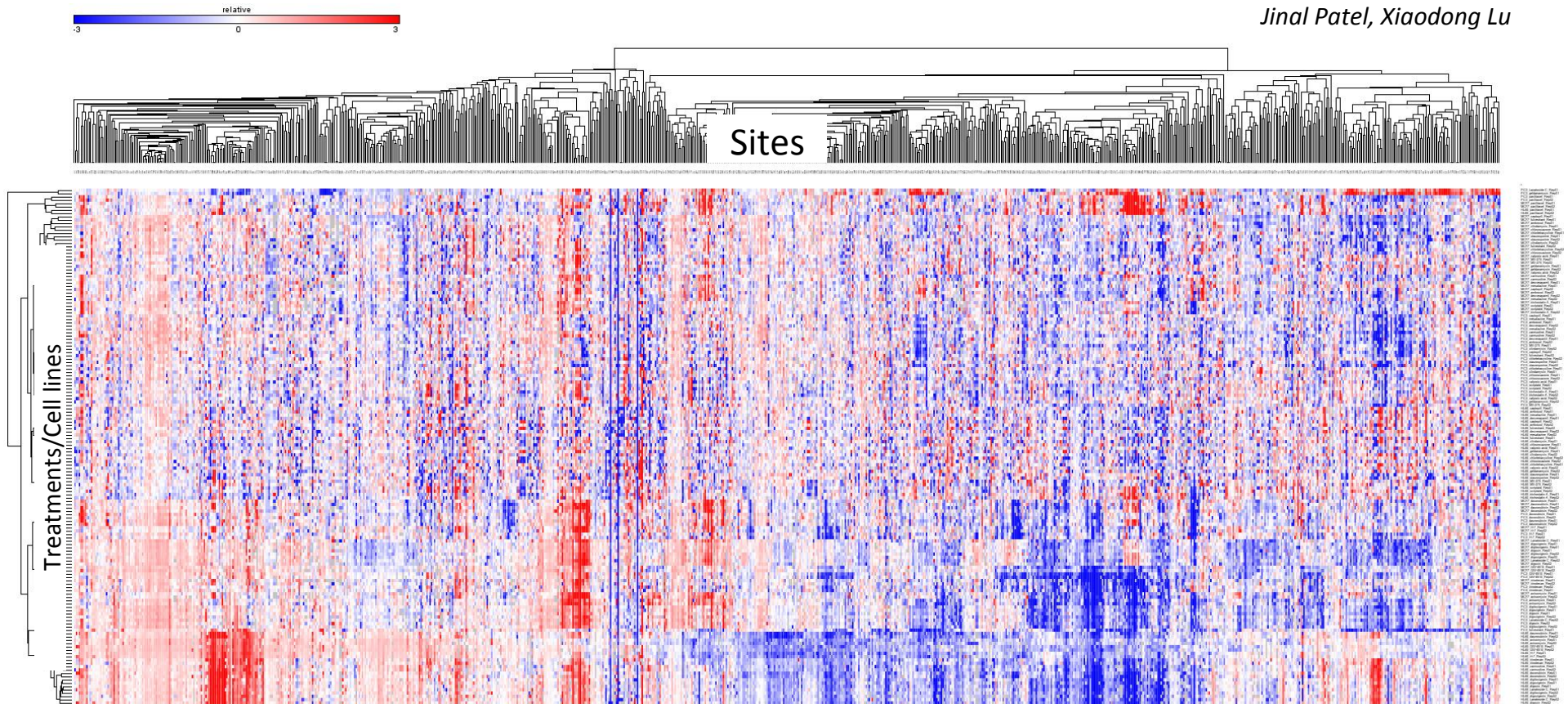
Learn a practical reduced probe set from the data



- Natural synergy between L1000 and P100 projects
- Exploit existing robust methods

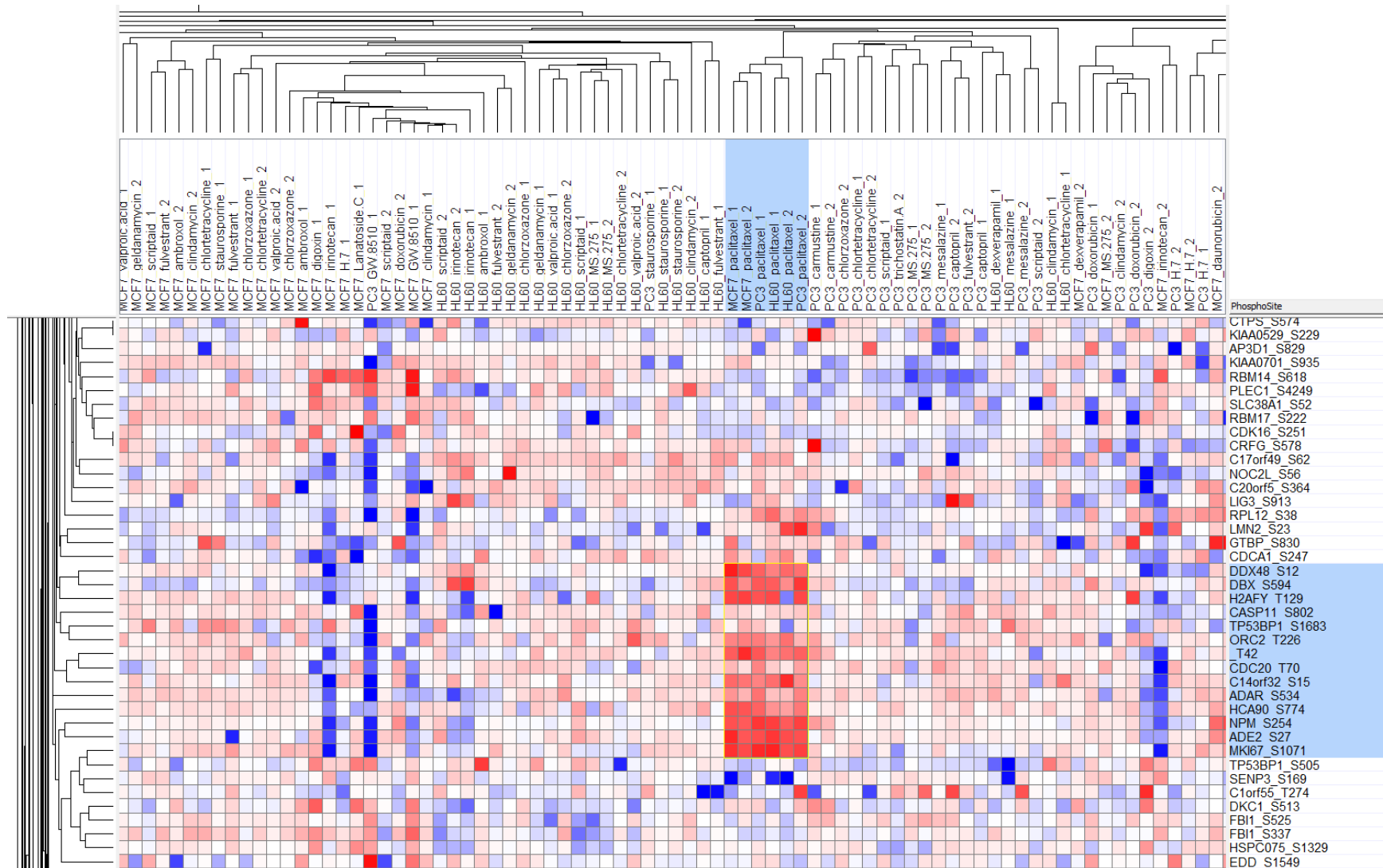
P100 Source Dataset

Jinal Patel, Xiaodong Lu



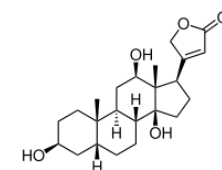
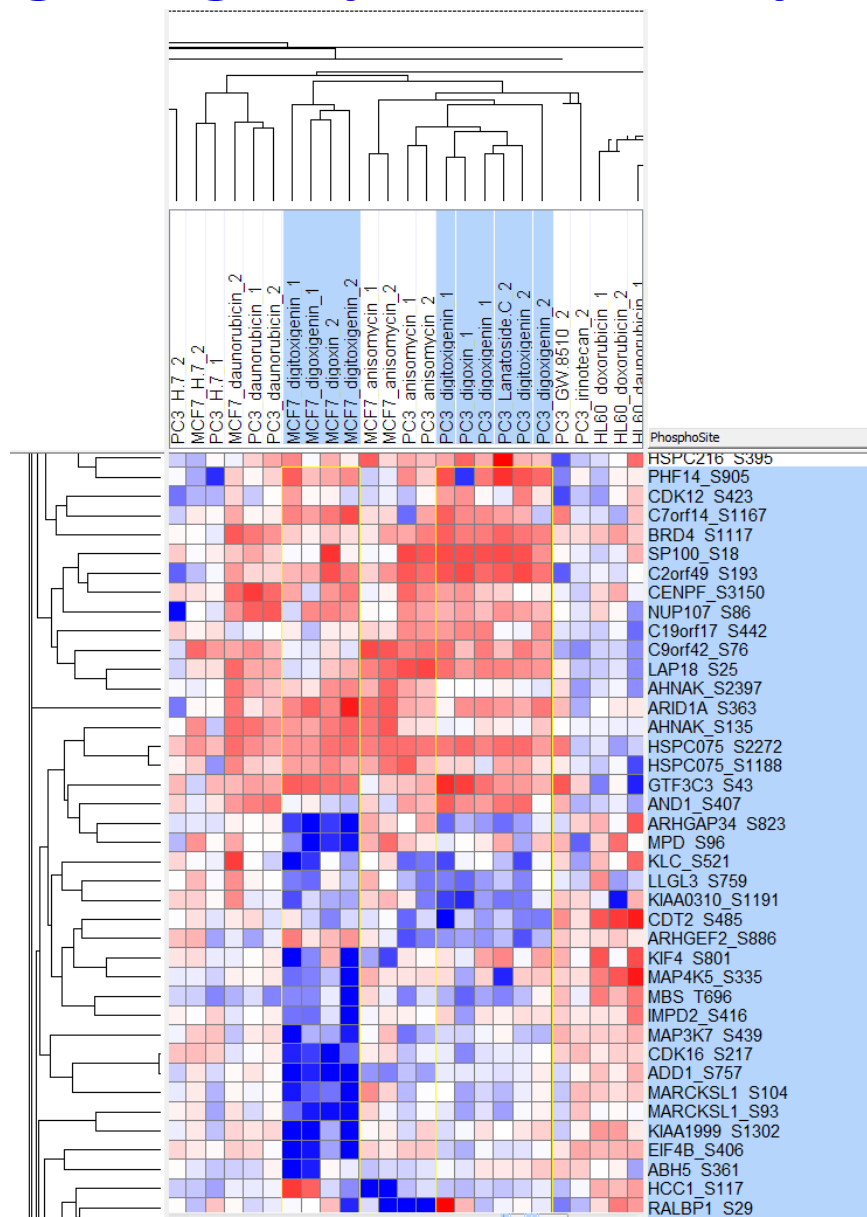
- Largest systematic set of perturbations with phosphoproteomics readout in existence
 - Over 10,000 phosphosites observed
- Over 1,200 sites present in >75% of all experiments

Signals span lineages

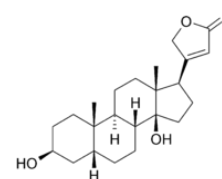


- All 6 Taxol treatments cluster – regardless of lineage

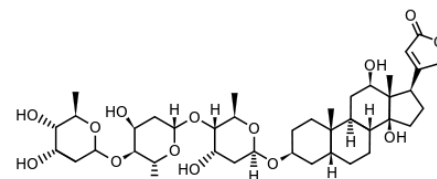
Signals group related compounds and replicates



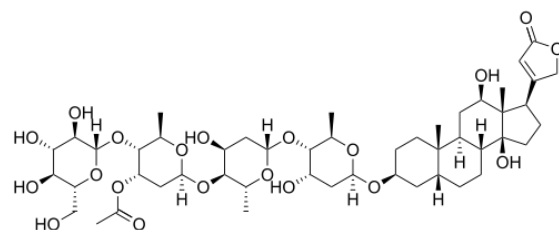
Digoxigenin



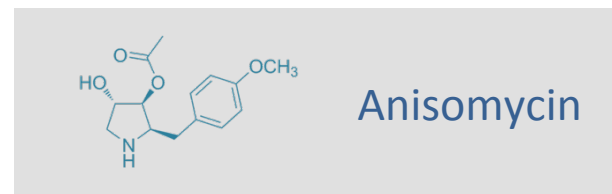
Digitoxigenin



Digoxin

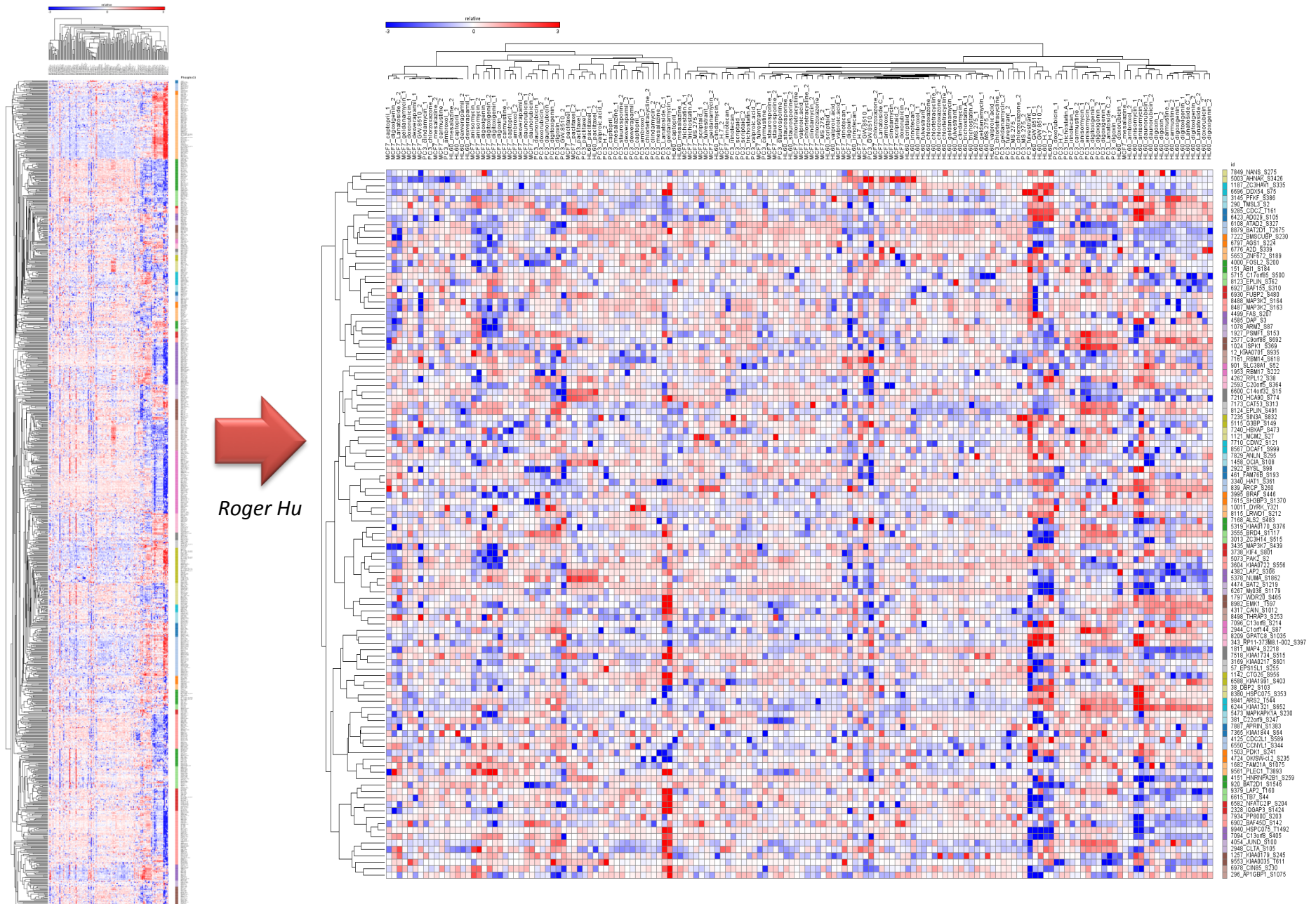


Lanatoside C



Anisomycin

Dimensionality reduction: >1000 to 55 with 2x probe redundancy



P100 Progress...

- Reduced Representation Targeted Phosphoproteomics Assay
- Large data set collected
 - Available at LINCS website
- Internal standards in hand
- Final assay configuration in progress
 - Expected to be deployable in multiple laboratories

Example: A targeted chromatin modification assay

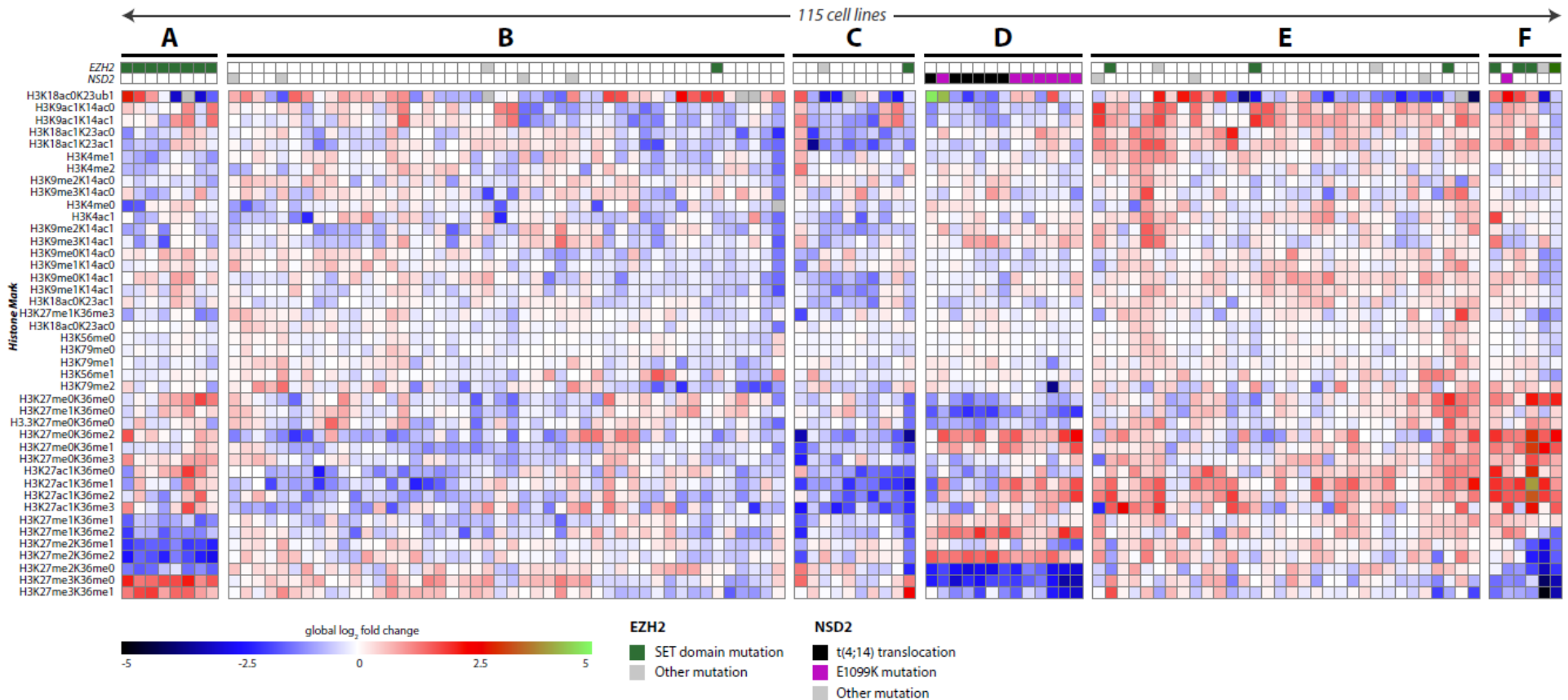
- Targeted high resolution assay against chromatin modification states
 - Over 60 possible combinations of histone marks are monitored
- We can query chromatin signatures from the MRM assay and infer function
 - Compare laboratory perturbations
 - Compare differing genotypes
 - Compare compound treatments
 - Compare across these experiment types
- The “CMap” of chromatin-space

SUV39H1 knockdown
signature
K9me2,me3 down
S10ph up

H3K4me0 - B110003
H3K4me1 - B110004
H3K4me2 - B110005
H3K4me3 - B110006
H3K9me0K14ac0 - B110008
H3K9me1K14ac0 - B110009
H3K9me2K14ac0 - B110010
H3K9me3K14ac0 - B110011
H3K9ac1K14ac0 - B110012
H3K9me0K14ac1 - B110013
H3K9me1K14ac1 - B110014
H3K9me2K14ac1 - B110015
H3K9me3K14ac1 - B110016
H3K9ac1K14ac1 - B110017
H3K9me0S10ph1K14ac0 - B110018
H3K9me1S10ph1K14ac0 - B110019
H3K9me2S10ph1K14ac0 - B110020
H3K9me3S10ph1K14ac0 - B110021
H3K9ac1S10ph1K14ac0 - B110022
H3K9me0S10ph1K14ac1 - B110023
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H3K9ac1S10ph1K14ac1 - B110027
H3K18ac0K23ac0 - B110028
H3K18ac1K23ac0 - B110029
H3K18ac0K23ac1 - B110030
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H3K18ub1K23ac0 - B110032
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H3K27me0K36me0 - B110034
H3K27me1K36me0 - B110035
H3K27me1K36me1 - B110036
H3K27me1K36me2 - B110037
H3K27me1K36me3 - B110038
H3K27me2K36me0 - B110039
H3K27me2K36me1 - B110040
H3K27me2K36me2 - B110041
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H3K27me3K36me1 - B110044
H3K27me3K36me2 - B110045
H3K27me3K36me3 - B110046
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H3K56me0 - B110054
H3K56ac1 - B110058
H3K56ub1 - B110059
H3K79me0 - B110060
H3K79me1 - B110061
H3K79me2 - B110062

Column cluster complete linkage by Pearson correlation

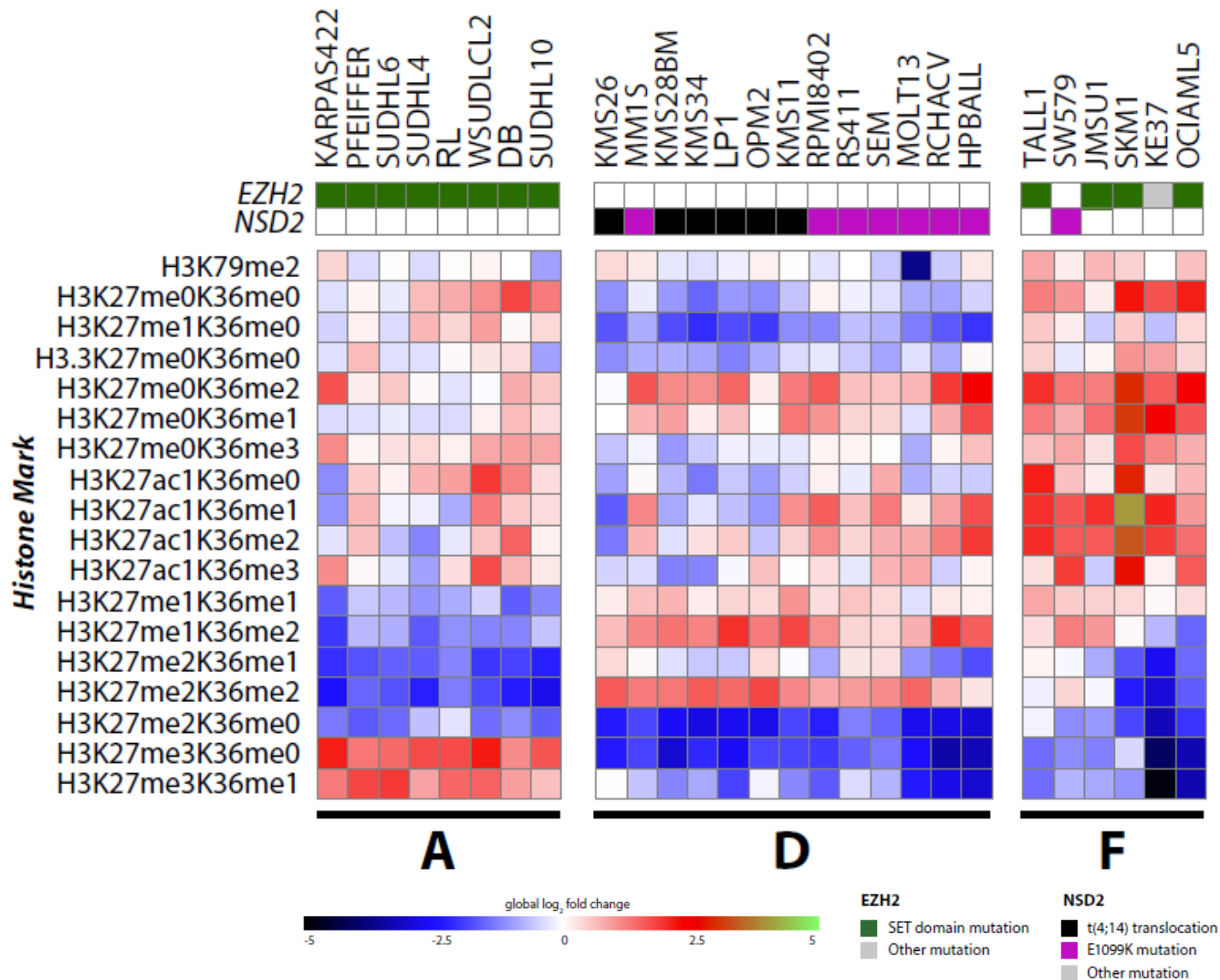
Molecular Chromatin Signatures of Genotypes



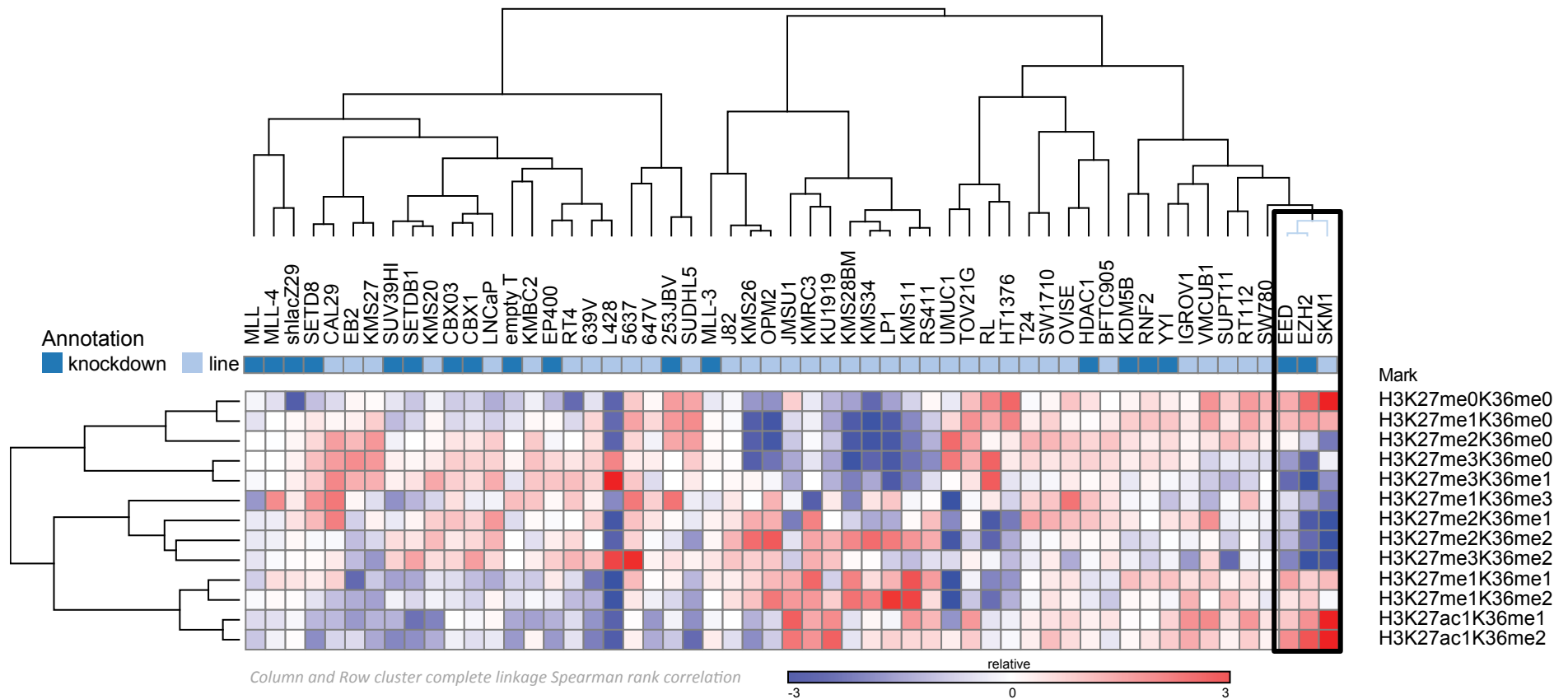
- Steady-state chromatin profiles of cell line in the Cancer Cell Line Encyclopedia

Jordan Taylor, CCLE team

Molecular Chromatin Signatures of Genotypes



Queries across data types in chromatin space



- Can co-cluster perturbation signature and mutation signature
- EZH2 mutation in SKM1 behaves like loss of function

Conclusions and Thoughts

- Proteomics must adapt to play nicely with functional genomics
- Major challenge is in generating (un)hole-y data at scale and cost
- Targeted proteomics offers a potential solution
 - Goal: universal cross-sample queries via standardization
- We are building a targeted phosphosignaling assay
- Targeted chromatin modification assay has already yielded promising results
- End-of-days for hole-y proteomics data!

Acknowledgements

- LINCS Program and Program Officers
 - U01 CA164186-01/Jaffe
- MacCoss Lab, Univ. of Washington
 - **Mike MacCoss** – co-PI
 - Brendan MacLean
- Broad Institute Proteomics Platform
 - Philipp Mertins
 - **Jinal Patel**
 - Steve Carr
- Broad Institute LINCS Centers
 - Todd Golub
 - Aravind Subramanian
 - **Xiaodong Lu**
 - **Roger Hu**
- CCLE Project Team
 - **Jordan Taylor** (Broad Proteomics)
 - Frank Stegmeier (Novartis)
 - HoMan Chan (Novartis)
 - Ellen Gelfand
 - Greg Kryukov
 - Levi Garraway
- Target Accelerator Team
 - Xiaoyun Wu
 - Yashaswi Shrestha
 - Jesse Boehm